

# The potential for bioelectrochemical detection of fecal contamination indicator organisms in environmental water

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*Ingestion of and contact with fecally contaminated environmental waters can cause both enteric and skin diseases. Therefore, many countries have adapted WHO Guidelines for Drinking-water Quality [1] and monitor drinking water supplies and recreational waters for such disease causing microbes [2]. Enteric bacteria, such as E.coli and other coliforms are commonly cultured from surface waters [3] and thus make good proxy organisms for fecal contamination. As the nutrient conditions of environmental waters differs from those in the gut, facultative anaerobes, such as E. coli, are more likely to stay viable than obligate anaerobes when shed from the mammalian gut. Methods like multiple tube fermentation (MTF) [4] and membrane filtration techniques (MFT) [5] were first used to exploit this principle and were developed as a method to determine the number of coliforms in an environmental water sample, and are still considered standard methods in water quality monitoring [6].*

*Although methods such as MTF and MPN are able to determine the presence of coliforms in environmental water samples, subsequent tests are required to obtain definitive confirmation of fecal contamination in a water sample. This can take up to an additional 72 hours [7]. In the last 30 years, several single step methods have been developed to detect fecal contamination of environmental waters using E.coli as a microorganism representative of fecal contamination. These methods are based either on detection of the presence of uidA gene by PCR [8], or by the ability of  $\beta$ -Glucuronidase, the expressed product of uidA in E.coli, to hydrolyse a substrate conjugated to D-glucuronic acid to elicit a fluorescence, colorimetric [7] or amperometric signal [9, 10].*

*We demonstrate that E.coli  $\beta$ -Glucuronidase is able to hydrolyse a novel glycoconjugate to give a signal that is detectable amperometrically by electrochemical methods. Furthermore, the onset of the amperometric signal is directly proportional to the initial number of viable E. coli in the sample. The reversible redox chemistry of this glycoconjugate means that it can be applied in a non-culture based self-sustaining bioelectrochemical system that would be compatible with automated remote detection. We discuss the results in the context of its potential to speed up the detection of E. coli, and other fecal indicator organisms, in the context of environmental water quality monitoring along with the economic implications.*

**Keywords**—E.coli, fecal contamination, detection, bioelectrochemical

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